

Slide Session  
1988 ASM ANNUAL MEETING  
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An Automatable, Colorimetric DNA Hybridization Test for *M. tuberculosis* Confirmation, BRAKEL, C.L., DONEGAN, J.J., LINN, C-I.P., MOLINA, M., POLICE, M.A., WANG, Z., and YANG, H. L. ENZO Biochem Inc., New York, N.Y.

An oligonucleotide-based DNA hybridization test for confirmation of *M. tuberculosis* (MTB) cultures has been developed that is amenable to either partial or complete automation. Following lysis (10 min.) of cultured specimens, the hybridization is carried out in two steps and can be accomplished in less than 2 hours (20-30 minutes "hands on" time), even when as many as 30-60 specimens are to be analyzed. The lysed cultured specimens are first hybridized against one modified oligomeric probe in solution and are then allowed to hybridize to a second probe coated onto wells of microtiter (ELISA) plates. After hybridization and washing, the hybrids are detected with streptavidin-biotinylated horseradish peroxidase. Signal is generated by enzymatic conversion of hydrogen peroxide and  $\alpha$ -phenylenediamine. Results can be read by eye, or quantitated with an ordinary ELISA photometer. To date the test has been 100% sensitive and specific. In a blind confirmation of 86 clinical isolates, 64 were correctly identified as MTB and 22 as non-MTB. In addition, 83 different species of bacteria, including 22 species of mycobacteria have been identified correctly as non-MTB. This methodology is suitable for the automated confirmation of any cultured organism provided suitable probes are available.

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Category designation

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An Automatable, Colorimetric  
DNA Hybridization Test for  
*M. tuberculosis* Confirmation

Special Thanks to

Jim Donegan

Patsy Lin

Margarita Molina

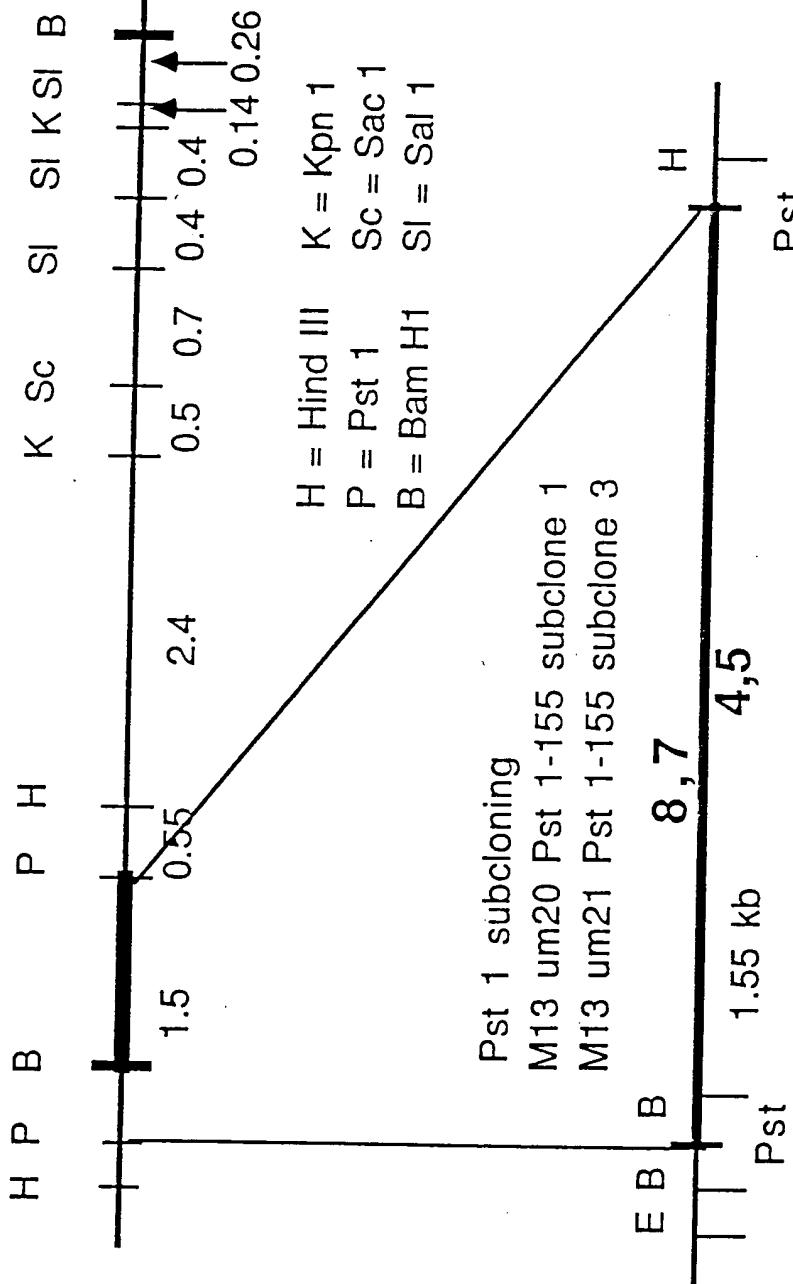
Marjorie Pollice

Zwang Wang

Huey Lang Yang

## Map and Sequences from MTB probe p24861

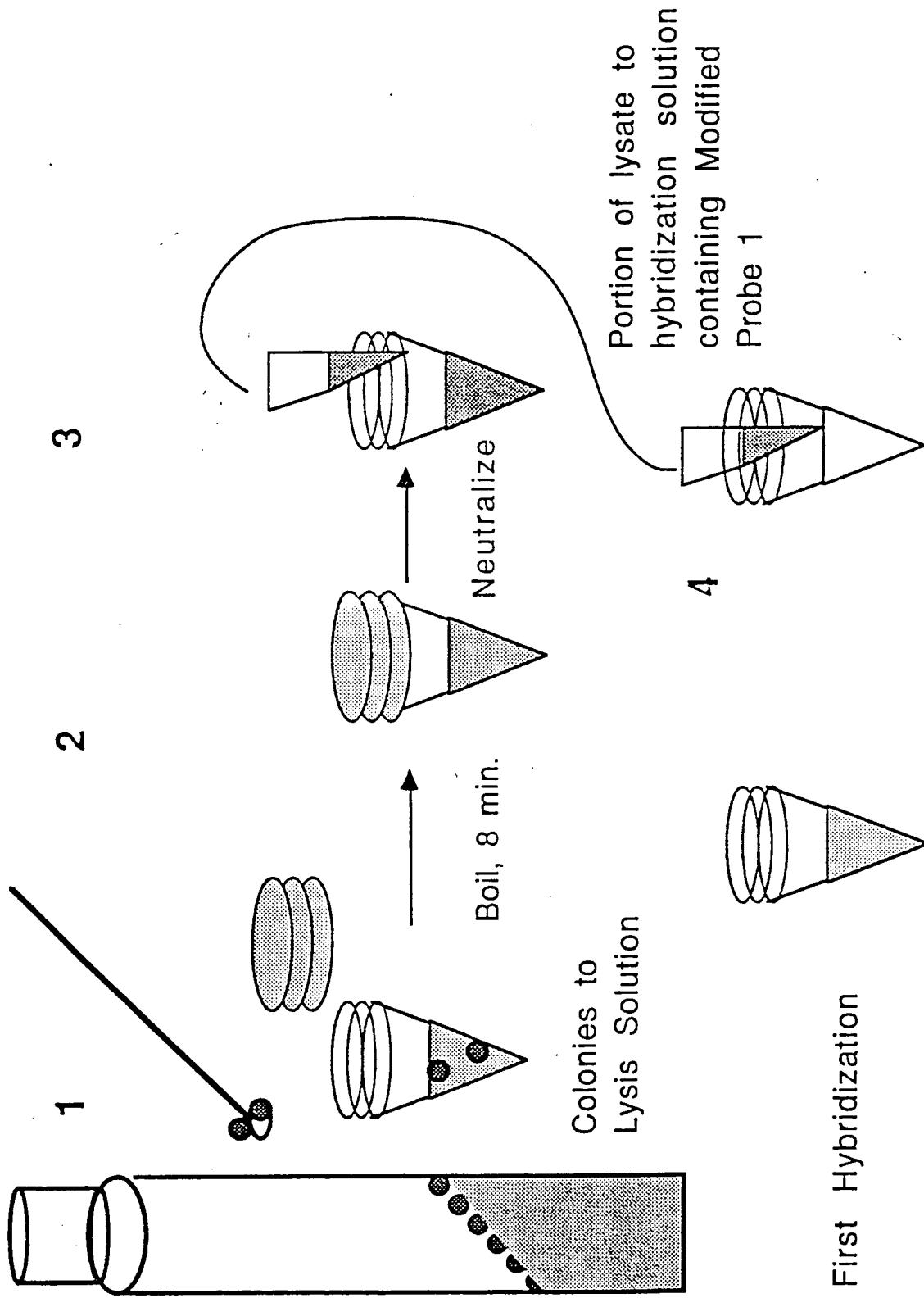
p24861 is a 7 kb insert in the Bam H1 site of pIB176, grown in HB101



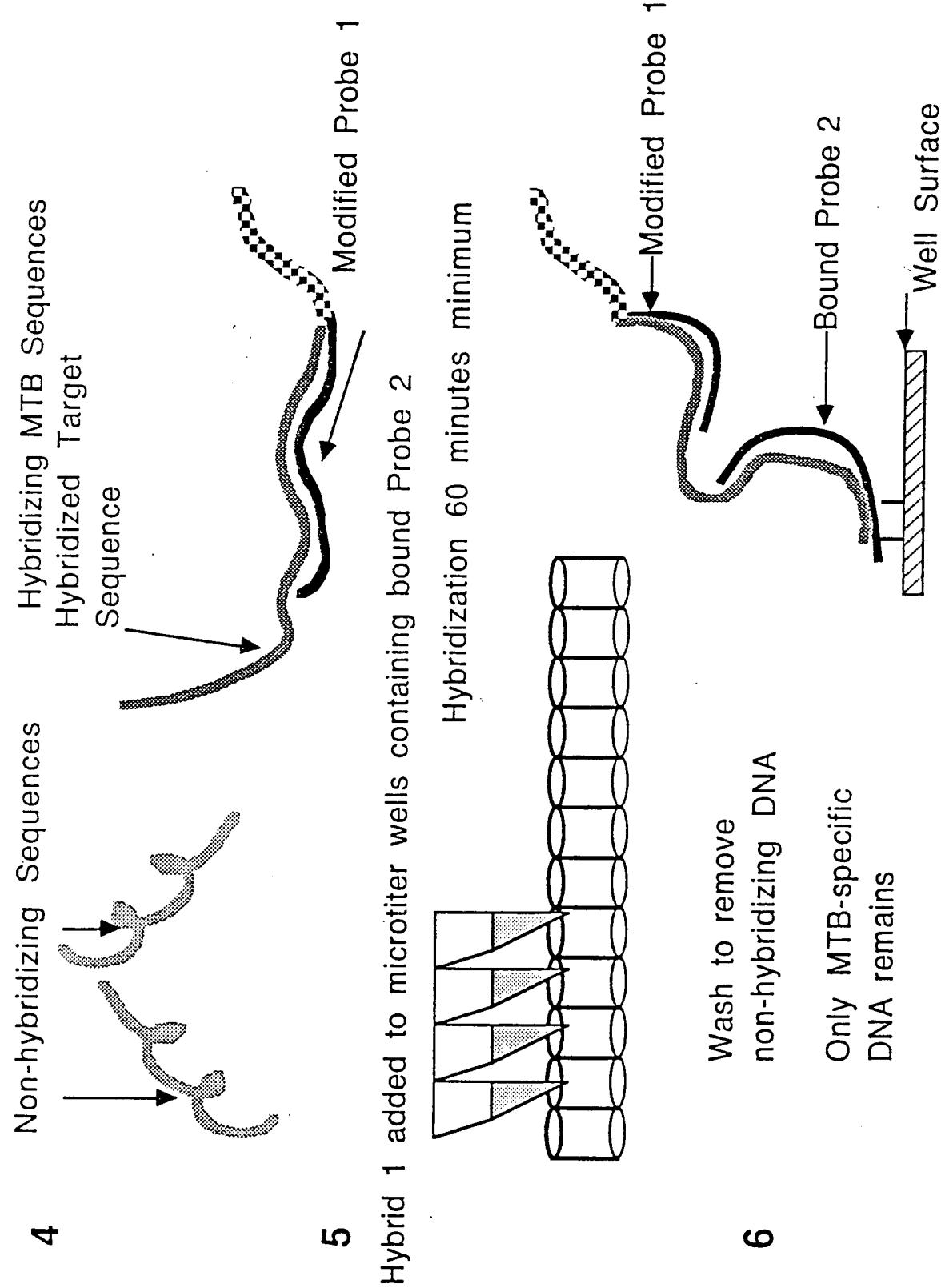
Sequenced by Dideoxynucleotide Methods

Oligonucleotides 4, 5, 7 and 8 are 30 base sequences synthesized using an Applied Biosystems Synthesizer. The two pairs are from opposite strands of the DNA.

## Specimen Lysis and First Hybridization

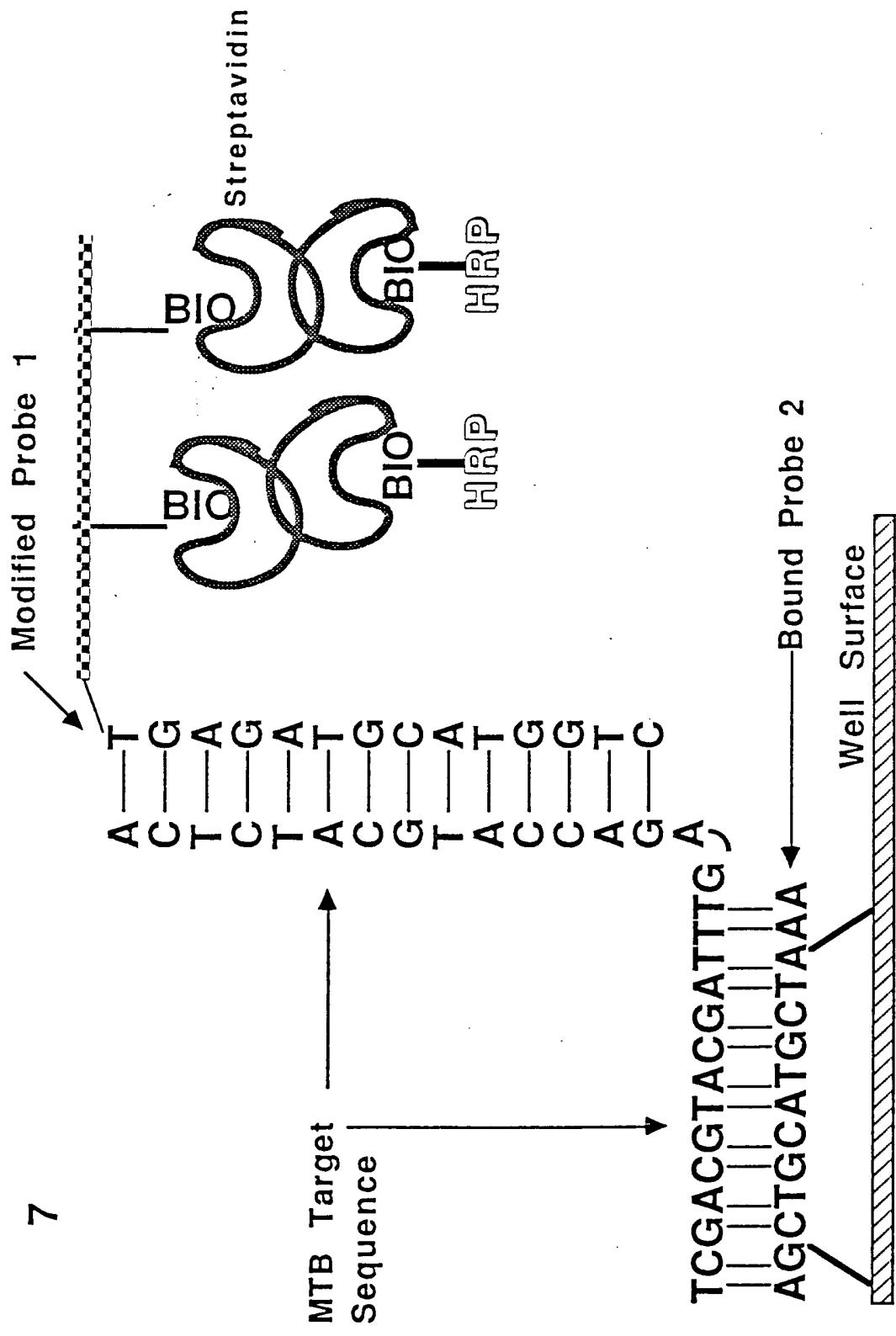


## Second Hybridization and Wash



## Detection of Hybridized MTB DNA

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(NOTE: Sequences are examples only, and are not the actual sequences used.)

## Cross Reaction Studies

<i>Acinetobacter calcoaceticus</i>	<i>Enterobacter aerogenes</i>	<i>R. sputi</i>
<i>A. lwoffii</i>	<i>Escherichia coli</i>	<i>Rhodospirillum rubrum</i>
<i>Actinomadura madurae</i>	<i>Fusobacterium nucleatum</i>	<i>Staphylococcus aurea</i>
<i>Actinoplanes italicus</i>	<i>Haemophilus influenzae</i>	<i>Streptococcus mitis</i>
<i>Arthrobacter oxydans</i>	<i>Klebsiella pneumoniae</i>	<i>S. pneumoniae</i>
<i>Bacillus subtilis</i>	<i>Legionella pneumophila</i>	<i>Vibrio parahaemolyticus</i>
<i>Bacterionema matruchotii</i>	<i>Microbacterium lacticum</i>	<i>Yersinia enterocolitica</i>
<i>Bacteroides fragilis</i>	<i>Mycoplasma hominis</i>	
<i>Branhamella Catarrhalis</i>	<i>M. pneumoniae</i>	
<i>Brevibacterium linens</i>	<i>Neisseria gonorrhoea</i>	
<i>Campylobacter jejuni</i>	<i>N. lactamica</i>	
<i>Chromobacterium violaceum</i>	<i>N. meningitidis</i>	
<i>Clostridium perfringens</i>	<i>Nocardia asteroides</i>	
<i>Corynebacterium aquaticum</i>	<i>N. brasiliensis</i>	
<i>C. diphtheriae</i>	<i>N. otitidis-caviarum</i>	
<i>C. genitalium</i>	<i>Nocardiopsis dassonvillei</i>	
<i>C. haemolyticum</i>	<i>Oerskovia turbata</i>	
<i>C. minutissimum</i>	<i>O. xanthineolytica</i>	
<i>C. pseudodiphtheriticum</i>	<i>Propionibacterium acnes</i>	
<i>C. pseudogenitalium</i>	<i>Proteus mirabilis</i>	
<i>C. pseudotuberculosis</i>	<i>Pseudomonas aeruginosa</i>	
<i>C. pyogenes</i>	<i>P. cepacia</i>	
<i>C. renale</i>	<i>Rahnella aquatilis</i>	
<i>C. striatum</i>	<i>Rhodococcus aichiensis</i>	
<i>C. xerosis</i>	<i>R. aurantiacus</i>	
<i>Deinococcus radiodurans</i>	<i>R. bronchialis</i>	
<i>Dermatophilus congolensis</i>	<i>R. chubaeensis</i>	
<i>Derxia gummosa</i>	<i>R. equi</i>	

## 63 Bacterial Species found to be Not Cross Reactive

**Cross Reaction Studies Mycobacterial Species**

**POSITIVE REACTIONS**

**MTB Complex**  
*Mycobacterium africanum*  
*Mycobacterium bovis*  
*Mycobacterium bovis BCG*  
*Mycobacterium tuberculosis*  
*Mycobacterium microti*

**Positive for MTB Complex**  
**Negative for 25 other**  
**Mycobacterial Species**

**NEGATIVE REACTIONS**

<i>M. asiaticum</i>	<i>M. kansasii</i>	<i>M. szulgai</i>
<i>M. avium</i>	<i>M. malmoense</i>	<i>M. terrae</i>
<i>M. chelonae</i>	<i>M. marinum</i>	<i>M. thermoresistibile</i>
<i>M. flavescens</i>	<i>M. nonchromogenicum</i>	<i>M. triviale</i>
<i>M. fortuitum</i>	<i>M. phlei</i>	<i>M. ulcerans</i>
<i>M. gastri</i>	<i>M. scrofulaceum</i>	<i>M. vaccae</i>
<i>M. gordonaiae</i>	<i>M. shimoidei</i>	<i>M. xenopi</i>
<i>M. haemophilum</i>	<i>M. simiae</i>	
<i>M. intracellulare</i>	<i>M. smegmatis</i>	

## Results of Testing

		Culture		Culture/GenProbe	
		+	-	+	-
Study 1	+	72	1 *	55	0
	-	1 * *	49	0	38
Study 2	+	DNA Assay		DNA Assay	
	-				

\*Originally identified as *M. xenopi*.  
Later found to contain MTB and  
*Corynebacterium pseudotuberculosis*.

All specimens were originally  
identified by use of the GenProbe  
MTB complex confirmation test.

\*\*Originally identified as MTB.  
Later found to be *Brevibacterium*  
linens.